## Pistoia The Allotrope 2023 Spring Connect Event

# LC-UV Methods and Results Interoperability using ADF

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- 25th April 2023

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#### Acknowledgements















+ The extended project team



## High Performance Liquid Chromatography (HPLC) instruments:











#### Pistoia Methods Database Project

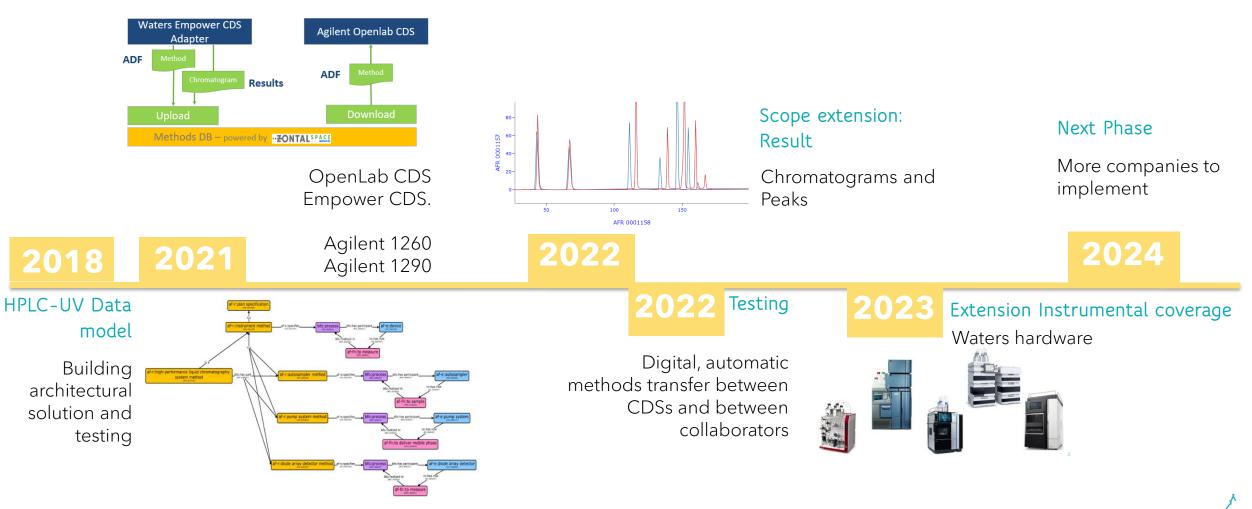
Pistoia chartered a project to transform analytical methods into standardized, machine-readable instructions that can be stored centrally and shared across different vendors / models of HPLC-UV to execute the methods



#### Methods Database Timeline





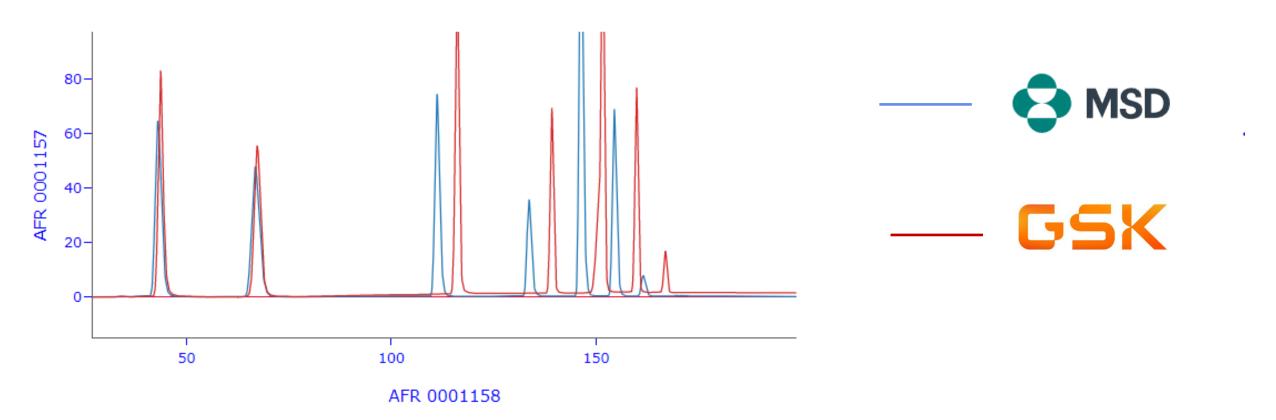


#### Methods DB Phase 1 Proof of Concept – Completed Transfer methods using OpenLab CDS through ZONTAL Cloud

Objectives	Import and export methods using OpenLab CDS Transfer methods across Agilent 1260 and 1290 instruments Overlay OpenLab CDS data on ZONTAL Cloud
Results	<ol> <li>Export and import gradient methods using OpenLab CDS</li> <li>Export and import isocratic methods using OpenLab CDS</li> <li>Transfer methods between Agilent 1260 and 1290</li> <li>Import third party method on OpenLab CDS</li> <li>Overlay Merck and GSK data</li> </ol>

#### Methods Portability across Companies: Comparison

Method's transfer on Agilent 1290





#### Methods DB Phase 2 Proof of Concept – Completed

### Transfer methods between Empower CDS and OpenLab CDS through ZONTAL Cloud and Orbis Gateway

Objectives	Import and export methods using Empower, OpenLab CDS and vice versa Transfer methods across Agilent 1260 and 1290 instruments Submit sample set from ZONTAL cloud to Empower CDS Overlay Empower and OpenLab data
Results	<ol> <li>Export and import gradient methods using Empower CDS</li> <li>Export and import isocratic methods using Empower CDS</li> <li>Transfer methods between Agilent 1260 and 1290</li> <li>Transfer methods between Empower and OpenLab CDS</li> <li>Import third party method on Empower CDS</li> <li>Overlay OpenLab and Empower data</li> <li>Sequence submission</li> </ol>

#### Methods DB Phase 2 Proof of Concept

#### Video Demo – Sequence Submission from ZONTAL to Empower CDS<sup>™</sup>

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	Mobile phase A		10% MeOH in 90% Wate	er	
	Mobile phase B		Acetonitrile		
	Flow rate		0.5 mL per minute (HPL0		Î.
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		0.75	90	10	
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		3.30	90	10	
		5.00	90	10	
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	Injection volume		5 microlitres		é de la companya de la
	Blank/Diluent		1:1 water/ methanol		
	Autosampler needle wash solvent		50/50 (v/v) water/ acetonit	trile	
	Seal Wash		10:90 IPA: Water		
	Data collection time/Run time		5 minutes		
	Number of samples		Blank + 1 sample		
	Sample concentration	St	andard 2 - Purchased from		
	Sequence Template		Blank (x3), Test mix (x6		
	Method File Names	AcqMet	-2800CCDS2_Agilent_1290 hod_RY801-B200-UPLC7_ od_RY818_C_208_Importer	TestMethod2	
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#### Methods DB Phase 2 Proof of Concept

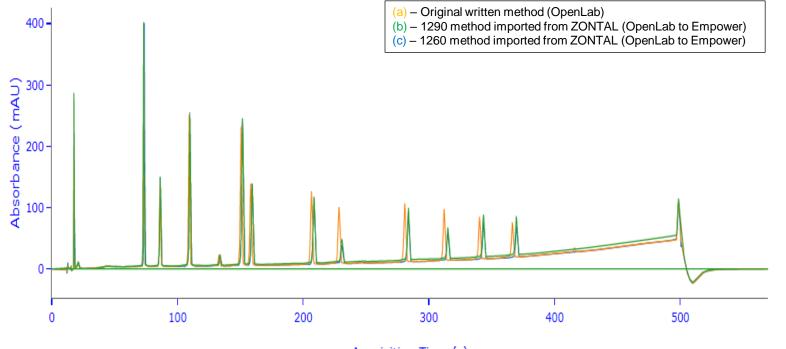
#### Video Demo – Empower and OpenLab Data Overlay on ZONTAL Space

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#### Phase 2 – Overlay of Empower and OpenLab CDS Data

Overlay of data collected on Agilent 1290 instrument using Empower and OpenLab CDS

The slight difference in peak height and retention time was expected as different lot of standards and mobile phases were used.



Acquisition Time (s)

Figure 2: HPLC chromatogram overlay of non-GMP 8-minute generic method (1290 Instrument): (a) original written method on 1290 using OpenLab CDS, (b) 1290 method imported from ZONTAL on Empower CDS and (c) 1260 method imported from ZONTAL on Empower CDS



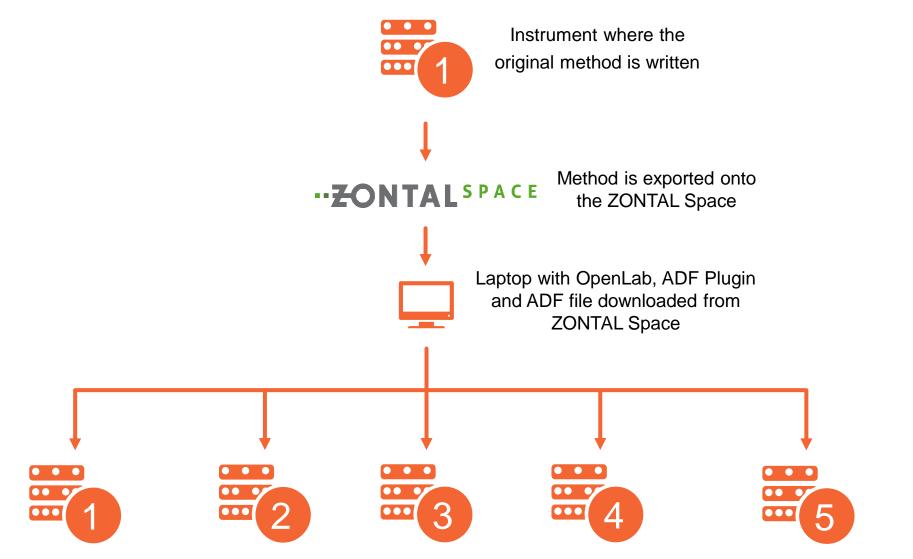
#### Methods DB Phase 3 Proof of Concept – Completed

Transfer methods using OpenLab CDS through the ZONTAL Cloud on a diverse set of instruments

Objectives	Import and export methods using OpenLab CDS Transfer methods across different Agilent Instruments (with different modules) Transfer methods from binary to quaternary pump
Results	<ol> <li>Install OpenLab and ADF plugin on laptop to facilitate instrument control</li> <li>Update the ADF plugin to support method transfer from binary to quaternary pump</li> <li>Export and import gradient methods using OpenLab CDS</li> <li>Transfer methods between Agilent 1290, 1260 and 1200 modules</li> </ol>
Next steps	<ul> <li>Repeat the experiments on two diverse instruments in the US</li> <li>Share findings internally and externally.</li> </ul>



#### 15 Minute Gradient Method Setup



ADF method files imported onto instruments with different modules stacking



#### HPLC/UPLC Instrument Configuration

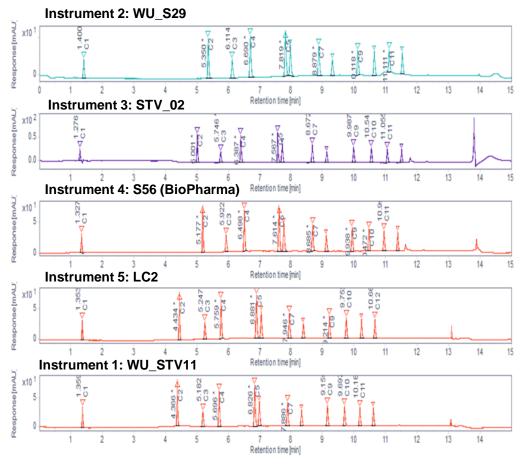
HPLC/UPLC Module Stacking

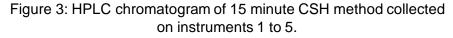
Module	Instrument 1: WU_STV11	Instrument 2: WU_S29	Instrument 3: STV_02	Instrument 4: S56 (Biopharma)	Instrument 5: LC2
Pump	G7120A: 1290 Infinity II Binary Pump	G1312A: 1200 Binary Pump	G7120A: 1290 Infinity II Binary Pump	G7111B: 1260 Infinity II Quaternary Pump	G1312B: 1260 Infinity Binary Pump
Injector	G7167B: 1290 Infinity II Multisampler	G1329A: 1100 Autosampler	G7167B: 1290 Infinity II Multisampler	G7167A: 1260 Infinity II Multisampler	G7167A: 1260 Infinity II Multisampler
Column Compartment	G7116B: 1290 Multicolumn Thermostats	G1316A: 1200 Thermostatted Column Compartment	G7116B: 1290 Multicolumn Thermostats	G7116B: 1290 Multicolumn Thermostats	G1316C: 1200 Thermostatted Column Compartment
Detector	G7117B: 1290 Infinity II Diode Array Detector	G1314B: 1200 Infinity Variable Wavelength Detector	G7117A: 1290 Infinity II Diode Array Detector	G7115A: 1260 Infinity II Diode Array Detector Wide Range (WR)	G4212B: 1260 Infinity Diode Array Detector



#### **15 Minute Gradient Method**

#### Results and discussion





- As shown in Figure 3, there the data collected on instrument 1 is comparable to instruments 2-5.
- The data obtain confirms our theory that the 1200/1260 has a lag in applying the gradient in comparison to the 1290 pump
- As expected, the same elution order was observed throughout all the 5 instruments.
- A slight change in difference in retention time was observed on various instruments but this was expected as each instrument will have a different dwell volume.
- The data proves that the ADF plugin can be used to transfer method across a wide range of Agilent HPLC/UPLC instruments.

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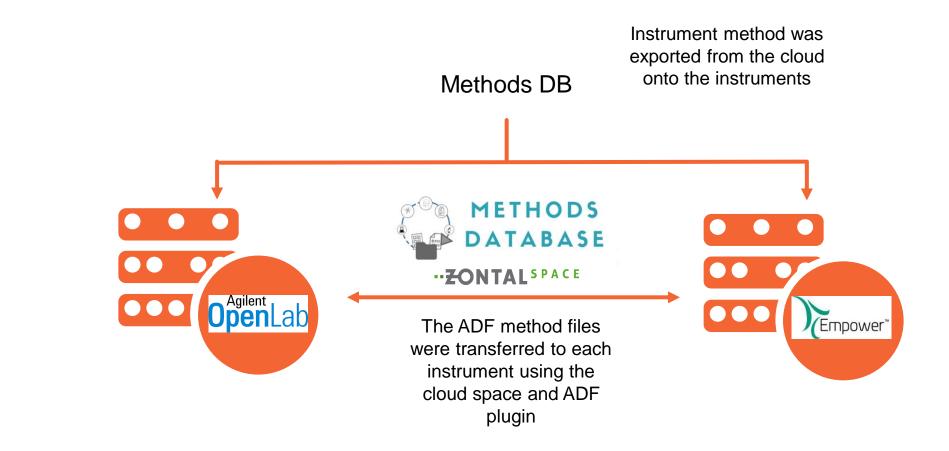
#### **Methods DB Phase 4 Proof of Concept – Completed**

Transfer methods between different manufacturers (hardware and CDS)

Objectives	Transfer methods between Waters and Agilent Hardware Transfer method between Empower CDS and OpenLab CDS
Results	<ol> <li>Transferred a gradient method between Waters Acquity and Agilent 1290 instrument (including autosampler temperature)</li> <li>Transferred a gradient method between Empower CDS and OpenLab CDS</li> <li>Overlay data acquired using both CDS</li> </ol>
Next steps	<ul> <li>Draft article to share results with analytical community</li> </ul>



#### Phase 4 Step Up



ADF files were shared between the two different manufacturers using the Zontal Space



#### Instrument Configuration

#### **UPLC Module Stacking**

Module	Agilent Hardware WU_STV11	Waters Hardware WU_STV0188
Pump	G7120A: 1290 Infinity II Binary Pump	Acquity UPLC H Class Quaternary Pump
Injector	G7167B: 1290 Infinity II Multisampler	Acquity UPLC H Class Sample Manager FTN
Column Compartment	G7116B: 1290 Multicolumn Thermostats	Acquity UPLC H Class Column Manager
Detector	G7117B: 1290 Infinity II Diode Array Detector	Acquity UPLC Photodiode Array Detector



#### **Diode Array Detector: Photodiode Array Detector**

		Genera La	
Signals	Advanced		λ Res
Acq         re         Wavelengen         Earownoor         Wavelength         Bandwidth           Signal A         Image: Constraint of the stress of		▼ 30.0 : to 400.0 : nm 20.0 : nm	Interp Use
Signal H         250.0 ;         4.0 ;         360.0 ;         100.0 ;         nm           Peakwidth         >0.025 min (0.5 s response time) (10 Hz)         •           Stoptime         Posttime	Attenuation: 1000   mAU  Margin for negative Absorbance  100  mAU  Autobalance	4 🔻 nm	DA [
As Pump/Injector     Off     1.00 ; min     1.00 ; min	✓ Prerun □ Postrun		Ch

#### **PDA Detector**

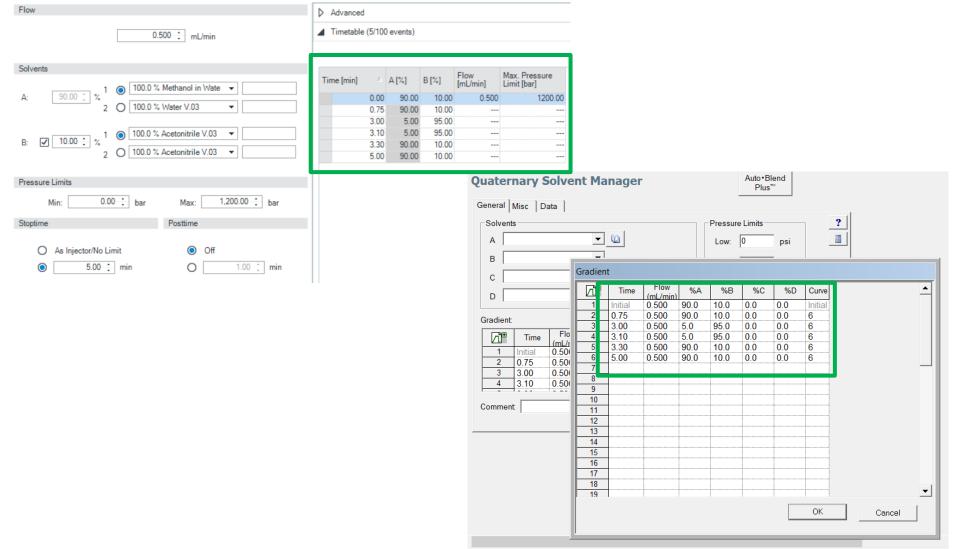
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▼ Enable 3D Data         λ Range:       191         191       nm         Resolution:       2.4         10       points/sec         Filter Time Constant       Exposure Time:         10       points/sec         Interpolate 2nd order filter Region         Use UV blocking filter (below 210nm)         Negative Absorbance Margin:         0.40         AU	eneral   2D Channels   Analo	og Out   Events	?
Resolution:       2.4 • nm         Sampling Rate:       Filter Time Constant       Exposure Time:         10 • points/sec       Fast • 0.1000 sec       Auto • msec         Interpolate 2nd order filter Region       Use UV blocking filter (below 210nm)       Negative Absorbance Margin:       -0.40 AU	▼ Enable 3D Data		
Sampling Rate:       Filter Time Constant       Exposure Time:         10 • points/sec       Fast • 0.1000 sec       Auto • msec         Interpolate 2nd order filter Region       Use UV blocking filter (below 210nm)       Negative Absorbance Margin:       0.40       AU	$\lambda$ Range:	191 nm to 400 nm	
10     points/sec     Fast     0.1000     sec     Auto     msec       Interpolate 2nd order filter Region       Use UV blocking filter (below 210nm)     Negative Absorbance Margin:     0.40     AU	Resolution:	2.4 • nm	
Use UV blocking filter (below 210nm) Negative Absorbance Margin: 0.40 AU			
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#### etector

General 2D Char	nnels Analog Out Events				
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🔽 Channel 2	Absorbance 💌	254	3.6	nm resolution	
Channel 3					
Compensation r	reference $\lambda$ Start 310	End	410 r	ım	

#### Binary Pump: Quaternary Pump





#### Column Compartment: Column Manager

Temperature		^ 🔺 A	Advanced			
Left:	Right:		able Analysis			
O Not Controlled	O Not Controlled		able Analysis when front door open			
● 40.0 ÷ °C	O 20.0 ¢ ℃		Left:		Right:	
O As Detector Call	O As Detector Cell		O With any temperature	O With any tempera		
O Unchanged	O Unchanged		When temperature is within	When temperature		
	Combined			• When temperatur		
			± 0.8 + °C for		± 0.8 ÷ °C for	
Valve Position/Column			0.0 📫 min		0.0 ‡ min	
O Use Current Column / Position		Valv	lve Position/Column After Run			
<ul> <li>Use Selected Column / Position</li> </ul>			_			
Position 1	- (J		Do not switch			
		0	Switch to position / column at beginning of run	Column Manager		
		0	O Increase valve position / column			
	0000	0	Decrease valve position / column	General Data		
		0	Use valve position / column			0
			Position 1	- Temperature		?
Enforce column for run				Column	Alarm Band:	
	~			<b>40.0 ▼</b> °c	± 1.0 °C	
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				Active Preneater.		
				Column Selection		
				valve Fosition.	Equilibration Time	
				Column 1 🗨	0.1 min	
				Comment		
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#### Multi sampler: Sample Manager

		The second se	-
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Injection volume: 5.00 🛟 µL	Eject Speea: Wait Time After Draw:	1.2 ; s	
	wait time Aiter Draw.	1.2 , 5	
Needle Wash	Needle Height Position		
Standard Wash 👻	Offset:	0.0 🛟 mm	
Stoptime Posttime	Use Vial/Well	I Bottom Sensing	
Supume	High Throughput		
As Pump/No Limit     Off	Sample Flush-Out Factor:	5.0 🛟	
O 1.00 <sup>↑</sup> min O 1.00 <sup>↑</sup> min	Injection Valve to Bypass fo	or Delay Volume Reduction	
	Enable Overlapped Injection	n	
	When Sample is Flush	hed Out	
	After Period of Time		
	0.00 ‡ n	ninutes after injection	
	Thermostat		
	On On Off	25 : °C	
		Sample Manager FTN	
		General Data Dilution Events	
		Solvents	Temperature Control
		Wash Solvent Name:	Column: Alarm Band:
The injection volume in Empower is specified		Water 🔽 💟	Off ▼ °C
		Purge Solvent Name:	Sample:
in the sequence table.		Water	Off ▼ °C  ± 5.0 °C
		Pre-Inject Wash:	
The ADF method file can also contain the		0 sec	Loop Offline:
auto sampler temperature which can be			Automatic 👻 min
· ·		Post-Inject Wash:	,
transferred across both manufacturers.		6 sec	Load Ahead
			Active Preheater:
			Disabled
		Comment	
			X Advanced

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#### Phase 4 – Data from different vendors

Data Comparison on Zontal Space – Agilent and Waters Hardware

The difference in retention time between the two manufacturers is due the different dwell volumes of the instruments used to execute the method.

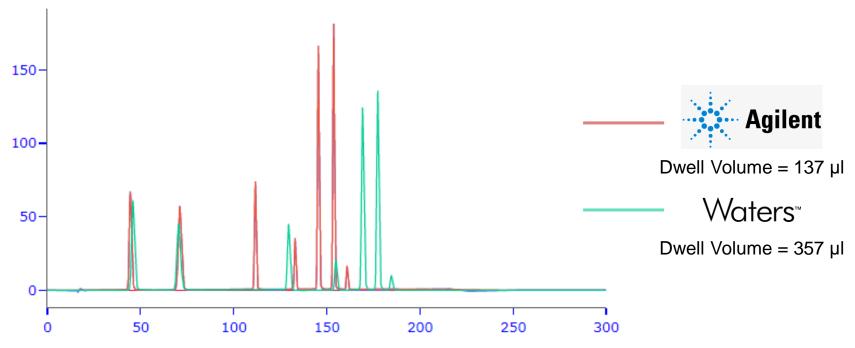


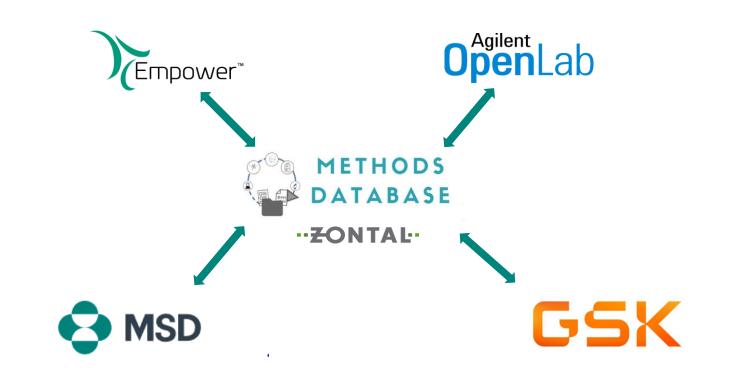
Figure 1: HPLC chromatogram comparison of the data collected Merck's gradient method on Agilent 1290 and Waters Acquity instruments at GSK.



### Conclusion

#### Method portability shown across

- ✓ Different instrument vendors
- ✓ Different chromatography data systems (CDS)
- ✓ Different pharmaceutical companies





#### How to get involved?



- Start implementation of the Methods Db
  - Lab/Instruments/Technical requirements
  - Interoperability across more hardware or software?
  - Own use case?
- Support the project/CoE

	NOW   6 MONTHS	NEXT   6+ MONTHS	
Next Phase of POC	<ul> <li>Define next phase to test a different dimension, perhaps pharma to CRO</li> <li>Documented plan for an implementable product</li> <li>Publish specifications for more software companies to test</li> <li>ROI calculator for Pharma companies</li> <li>Calculation of pharma investment</li> <li>Plan for sources of funding</li> </ul>	<ul> <li>Efforts to launch implementable product</li> <li>Additional software companies test in order to make product software agnostic</li> <li>Involve new instrument company</li> <li>Progress with columns + data analytics</li> </ul>	
Socialization	<ul> <li>Material plan to use broad-based marketing to socialize progress so far and upcoming plan</li> <li>Publish webinar series from the day</li> <li>Publish pharma test cases story in addition to June 2022 press release</li> <li>Strengthen pitch with ROI and investment data</li> <li>Bring together more pharma companies to join initiative</li> <li>Tap into support of new Pistoia member, Charles River</li> </ul>	<ul> <li>Bring together more pharma companies to join initiative</li> <li>Proliferation through organizations to warrant future development</li> <li>Gather contacts to target from Pistoia commercial partners</li> <li>Identify mutual customers across businesses represented to expand to</li> </ul>	









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### Any questions? Contact us: methodshub@pistoiaalliance.org

